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Ovine Enzootic Abortion (OEA): a comparison of antibody responses in vaccinated and naturally-infected swiss sheep over a two year period

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Abstract

Background

Prevention and control of ovine enzootic abortion (OEA) can be achieved by application of a live vaccine. In this study, five heifer flocks with different vaccination and infection status were serologically tested using a competitive enzyme-linked immunosorbent assay (cELISA) specific for *Chlamydia* (*Cp.*) *abortus* over a two-year period.

Results

Sheep in flock A with recent OEA history had high antibody levels after vaccination similar to flock C with natural *Cp. abortus* infections. In contrast, OEA seronegative heifers (Flock E) showed individual animal-specific immune reactions after vaccination. Antibody levels of vaccinated ewes in flock B ranged from negative to positive within three years after vaccination, respectively. Positive antibody values in the negative control flock D (without OEA or vaccination) are probably due to asymptomatic intestinal infections with *Cp. abortus*. Excretion of the attenuated strain of *Cp. abortus* did not help vaccination brought the ewes as not observed in vaccinated animals of Flock E.

Conclusions

The findings of this study indicate that, using serology, no distinction can be made between vaccinated and naturally infected heifers. As a result, confirmation of a negative OEA status in vaccinated animals by serology cannot be determined.

Background

Chlamydia abortus (formerly *Chlamydia psittaci* serotype 1) is the most common infectious bacterial cause of abortion in small ruminants in Switzerland with previous studies demonstrating that 39% of the examined abortions in sheep and 23% in goats were caused by this agent [1]. In the Swiss canton of Graubünden, a mountainous region in the eastern part of the country, the economic losses associated with ovine and caprine abortion (OEA) are significantly higher than in other cantons [2].

C. abortus is generally introduced into immunologically naïve flocks by a latently infected animal with the agent being subsequently transmitted from a aborting ewe or goat to a large number of infectious *Chlamydia* in the foetal membranes and placentas [3]. In newly infected flocks, up to 30% of the ewes may abort in the first trimester of gestation or give birth to weak or dead lambs. After abortion, the ewe in these flocks may develop protective immunity. Subsequent yearly losses in endemicity in infected flocks may decrease to a low level (e.g. 5–10%) with the ewe either aborting or not having a lamb in the following years likely to suffer abortions during her initial pregnancies [4,5].

Prevention and control of OEA is achieved by vaccination and/or treatment with oxytetracyclines [6]. Two vaccines against *Chlamydia* abortion are licensed in Switzerland by the Federal Veterinary Office (FVO) in Bern. The first of these available was a live, egg-grown, formalin-inactivated, whole-organism vaccine (OvaxChlamidia, Farnim, Italy) which reduces the incidence of abortion in vaccinated herds but not completely [7–10]. Since December 2002, a non-virulent, temperature-sensitive, live *Chlamydia* vaccine (Ovilis® Enzovax, Intervet, The Netherlands), which is marketed to induce a long-lasting protection, has been made commercially available in Switzerland. The heat-inactivated strain 1B, which forms the basis of this vaccine, was obtained from the virulent *C. abortus* strain A/B7b by nitrosoguanidine mutagenesis [11–13].

In 2005, as mallpi lots tudyw asunde rtakent ode terminei fa dministrationof va ccinest o protects heep flocks from OEA would result in antibody levels not high enough for complement-fixation test (CFT) and not high enough for competitive enzyme-linked immunosorbent assay (cELISA) test similar to those following natural infection [14]. A further vaccination with the inactivated vaccine (Ovax Clamidia) only showed a weak detectable antibody response. In contrast, vaccination with the attenuated live vaccine (Ovilis[®] Enzovax) resulted in a detectable antibody titer in all tested sheep.

The aim of this study is to investigate the serological response of sheep over a two-year period in the field to compare flock-level ELISA responses between (a) vaccinated (live vaccine), (b) naturally infected and (c) non-infected sheep flocks. It was anticipated that the follow-up study of the humoral responses could possibly discriminate between vaccinated and naturally OEA-infected sheep. An additional objective of this study was to also attempt to detect chlamydiae and/or the attenuated strain of *Cp. abortus* in the live vaccine in conjunctival swabs of sheep.

Results

Serological results and abortion cases

cELISA classifications (frequency and proportion positive), median titer and respective range of positive classifications in flocks A, B, C, D and E over the four different investigation dates are shown in Table 1. The comparison between vaccinated and non-vaccinated animals in flock B and E is shown in Table 2. Figure 1 shows the interranges (boxplots) of all examined sheep in the live flock over the four investigations.

All ewes (n=15) of Flock A were serologically positive after vaccinations showing a high median antibody value of 91.7%. The median antibody level of positives sheep (n=13)

decreased marginally to 86.6% in autumn 2005. In spring 2006 and autumn 2006, the seroprevalence in the flock was 73% (n=11). The median antibody value of the positives heep was 81.3% (spring 2006) and 82.3% (autumn 2006).

In spring 2005, two years after the first vaccination, six out of 14 vaccinated heep in flock B had a positive serological result (median antibody value 62.9%), whereas two out of 12 non-vaccinated heep in the flock were positive. The number of positives heep decreased to three and two in the vaccinated group (n=14) in autumn 2005 and spring 2006, respectively. In the non-vaccinated group, one sheep tested positive in autumn 2005, but none in spring 2006. In autumn 2006, the number of positives heep increased in the vaccinated (n=6) and non-vaccinated (n=2) group, although a few abortions were not reported. The median antibody values in the two groups were comparable, but the values were slightly greater than 60%.

Flock C (naturally infected flock) served as the positive control. The seroprevalence in heep in spring 2005 was as high as 82% (n=14). The median antibody value in the positive group was 82.9%. The seroprevalence remained continuously high (76%-88%) during the whole study period and median antibody values in positives heep were above 70%. In autumn 2005, newborn lambs were largely negative and had a significantly lower median antibody value than older ewes (Kruskal-Wallis test, $p < 0.05$) (data not shown). The ewes with the confirmed chlamydial abortion in spring 2005 had positive antibody levels shortly after the abortion, during the period comparable to the other animals in the flock (50.5%-77%). The seroprevalence in goats after confirmed chlamydial abortion in the four animals in spring 2005 was 100% (n=4) with a high median antibody value of 91.6% (data not shown). In contrast to the heep, all goats remained serologically positive with very high antibody values (71.2%-97.5%) over the whole testing period (data not shown).

Flock D served as the negative control for this study. Despite this, 21% (n=13) of the ewes showed positive results in spring 2005, whereas 44% (n=28) of the ewes had a negative serological result and 35% (n=22) of the animals showed questionable readings. The median

antibody values of the positive animals were 69.5 %. Half a year later, in autumn 2005, 21 animals continued to be serologically positive. In spring 2006, the seroprevalence increased to 46%, whereas the mean antibody values of the positive animals were comparable to spring and autumn 2005 (around 69%). In autumn 2006, the number of serologically positive was decreased to 30% (n=19), whereas the mean antibody values of the positive heep increased to 74.3%.

Prior to vaccination in spring 2005, only one animal in Flock E was positive in the vaccination group (antibody value 61.4%), whereas 25 heep (50%) were positive in the non-vaccinated group (n=50). All 3 heep of the vaccinated group were serologically negative in autumn 2005 and therefore elected for vaccination in winter 2005. The non-vaccinated group showed seroprevalences between 38- 48% from autumn 2005 to 2006 and the median antibody values of the positive animals were consistently between 80.4 -82.2%. In comparison to vaccinated heep in Flock A, none of the animals vaccinated in winter 2005 were serologically positive in spring 2006. In autumn 2006, one of the vaccinated heep had an antibody value of 73.2%, whereas the other 12 vaccinated heep had negative (n=6) or questionable values (n=6).

Statistical comparison of mean titers

In flocks A (all animals vaccinated), C and D (no animals in both flocks vaccinated, Figure 1), differences in titer values between the sampling periods were always highly significant in the RMA NOVA model ($p < 0.01$). In Flock B, with vaccination in the two sampling periods 2 and 3, both vaccinations status and the interaction term between vaccination and site were statistically significant ($p < 0.05$). In Flock E, in which vaccination took place before the first sampling, both main effects were significant (time: $p < 0.05$; vaccination status: $p < 0.01$), while the interaction term was not.

PCR results of eye swabs

In FlockE, 118 conjunctival swabs were collected before application of the live vaccine in autumn 2005. No obvious signs of ocular surface diseases such as conjunctivitis and keratitis were observed in any animal. IGS-SP CRs screening detected 22 samples that were positive for chlamydial DNA. Sequencing of these PCR products identified 18 samples that had a greater than 98% sequence similarity to *Cp. abortus* [GenBank:CR848038.1]. One sample each was revealed to be positive for *Cp. pecorum* [GenBank:CPU68434] and *Cp. felis* [GenBank:AP006861.1]. The identity of the two samples could not be determined.

Five months after vaccination, in spring 2006, 118 eye swabs were sampled in the same flock. 12 samples were tested positive by the IGS-SP CR but a low level of non-vaccinated ewes. Of these samples, 5/12 were positive for *Cp. abortus* [GenBank:CR848038.1] while three were positive for *Cp. pecorum* [GenBank:CPU68434]. The identity of the four samples could not be determined. None of the vaccinated sheep showed a positive IGS-SP CR result, thus it was concluded that no excretion of the vaccine strains had occurred.

Discussion

This study represents the first long-term chlamydial serological study comparing vaccinated and non-vaccinated flocks in Switzerland. The investigations were undertaken in the Canton Graubünden, where numerous chlamydial abortions in sheep were previously reported [1] and the high prevalence (43%) of *Cp. abortus* in Swiss cantons was observed [2]. The results obtained from this study confirm the previous observations of the pilot study [14] that serology (cELISA) cannot be used to distinguish between vaccinated and live attenuated vaccine and naturally-infected sheep. The antibody titer is not hereditarily vaccinated Flock A was comparable to Flock C in which a cut-off infection of *Cp. abortus* occurred at the same time. In Flock A, very high antibody levels (around 90%) were visible in

every vaccinated sheep (n=15), whereas antibody levels of sheep not vaccinated were somewhat lower (around 60%) 21 days post vaccination [14]. A serological abortion was reported in Flock A in the heparst, sheep could have been already serologically positive before vaccination and they might have had antibody levels could represent a novel flock abortion associated antibody levels. These antibody levels of positive animals decreased in both flocks (A and C) from spring 2005 to spring 2006. A serological abortion was diagnosed in one goat from Flock C in autumn 2006 explaining the increasing seroprevalence and antibody levels in this group of animals at that time. These antibody values in the goats of Flock C after a natural infection with *Cp. abortus* were higher and persisted at a very high level (80 to 90%) over the observation period compared to the situation in sheep. No correlation with protection was seen however as a serological abortion occurred in a seropositive goat which had previously aborted. This observation also indicates that not her goats in flock C are not infected (R. Thoma, personal communication). Goats treated with the live vaccine also aborted. In general, it is known that if *Chlamydiae* are introduced in a naive flock, the losses are much higher in goats (60%) than in sheep (30%). The differences between goats and sheep are consistent with previous records and therefore remain unexplained [15,16].

Antibody levels of vaccinated ewes of Flock B ranged from negative to positive within three years after vaccination, respectively. Questionable antibody levels are either attributed to undiagnosed *Cp. pecorum* infections [17] or are possibly due to the vaccination in spring 2003. In a similar situation of the naturally infected sheep (Flock C), as low decrease of antibody levels was observed over the sampling period. These observations strongly suggest that serology (cELISA) cannot be used to distinguish between vaccinated and the live attenuated vaccine and naturally-infected sheep as anticipated in the previous study [14]. As a direct consequence of this, the confirmation of negative O EAs in vaccinated animals by serology cannot be made. This is unfortunately a reliable confirmation is important

if a naïve batement of OEA through assembly of OEA-free flock is to be performed as undertaken by the sheep and goat health schemes in England and Wales and the Premium Health Scheme in Scotland.

Positive antibody values have been observed in the negative control flock (Flock D), which had not been vaccinated and was free from chlamydial abortion. A new explanation for the observations of an increasing antibody value amongst his flock is that the animals may have asymptomatic intestinal infections with *Cp. abortus* as presumed in previous studies [17,19]. An alternative scenario is that the ewes were infected with a less virulent strain of *Cp. abortus*, which provoked seroconversion but not abortion [17,20]. Fluctuations in the antibody levels could be the result of bacterial shedding during estrus which provokes an induction of antibody levels without causing abortion [21,22]. Unfortunately, little is still known about this time about the ability of *Cp. abortus* to persist in animals (and the anatomical location of the persistent infection) compared to other chlamydial species, which require more investigations.

In Flock E, the serological reaction of 13 selected vaccinated sheep and the 50 non-vaccinated sheep in the flock was evaluated. Surprisingly and in contrast to the observations in the previous pilot study [14] and in the two vaccinated flocks A and B, six of 13 vaccinated sheep of Flock E showed no seroconversion eight months after vaccination. Only once we had a positive serological result (73.2%), comparable to the vaccinated sheep of Flock A and the naturally OEA-infected sheep of Flock C. The remaining six ewes had questionable antibody levels. The primary difference between animals in flocks A and E was the high variability of antibody levels in vaccinated animals. These results suggest that individual immune reactions between sheep can vary considerably.

Sampling of conjunctival swabs from sheep in Flock E was performed to detect and compare the presence of chlamydial DNA before and after vaccination. Furthermore, a possible

excretion of the vaccine brought here you could be screened with this approach. Although chlamydia were frequently detected by PCR in conjunctival swabs of sheep, the attenuated strain of *Cp. abortus* did not have vaccine was not detected in swabs collected from vaccinated sheep. The incidence of *Cp. abortus* and *Cp. pecorum* and even *C. suis* in clinically healthy non-vaccinated sheep was previously observed in a recent study [23]. The significance of this possible mode of transmission for OEA needs further investigation.

Conclusions

The findings of this study strongly suggest that serology (cELISA) cannot be used to distinguish between vaccinated and healthy attenuated vaccine and naturally-infected sheep. The course of antibody levels, nevertheless, can vary between individual animals and flocks. Compared to sheep, goats displayed higher antibody levels, which persisted for a longer time period but do not correlate with protection. The attenuated strain of *Cp. abortus* did not have vaccine was not detected in eyes swabs collected from vaccinated sheep.

Methods

Flock details

Fifty different sheep flocks in theanton Graubünden were followed over a two-year period with four visits. These five flocks were available for the study in spring 2005 through an established collaboration with the tertiary authorities in theanton Graubünden.

Due to constant turnover in each flock (i.e. slaughter of old and sick ewes, birth of lambs, introduction of new animals) the number of animals tested during the study was small and the

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blood samples were centrifuged at 3000× g for 10 minutes and stored in Nunc CryoTubes (Nalge Nunc International, Roskilde, Denmark) at -20°C until further processing.

cELISA

Serum samples were tested by the competitive enzyme-linked immunosorbent assay (cELISA) using the monoclonal antibody Ab188 directed against the variable regions 1 (VS1) and 2 (VS2) of the outer membrane protein (MOMP) of *Cp. abortus*, according to the protocol of Saiti-Montesano et al. [17]. The results of the ELISA were expressed as 'percentage of inhibition' corresponding to the antibody concentration in the sample.

Inhibition values above 55% were considered positive for infection with *Cp. abortus* (positive cut-off) whereas inhibition values between 30–55% were classified as questionable, attributable to either *Cp. abortus* or *Cp. pecorum*, a widely distributed chlamydial agent in small ruminants causing diseases such as arthritis/conjunctivitis and pneumonia syndrome in lambs and also subclinical intestinal infections [18,19]. Inhibition values below 30% were considered to be negative [17,24].

PCR of eye swabs

Conjunctival swabs (Cytobrushes, Berdat Charles, Bourroux, Switzerland) were collected from Flock E before and after vaccination of the investigated population for the excretion of chlamydiae and/or the *Cp. abortus* vaccine strain through the eye. Before application of the vaccine, conjunctival swabs from every sheep in the flock (n = 118) were collected in autumn 2005. Five months following vaccination (spring 2006), the second conjunctival swabs samples were taken from every sheep in the flock (n = 118). Cytobrushes were placed in 1.5-ml Eppendorf tubes and stored at -80°C until further processing. DNA extraction from all swabs was performed as described previously [25] using a commercial DNA extraction kit (DNeasy Tissue Kit[®], Qiagen, Hombrechtikon, Switzerland).

The conjunctival swabs were investigated for the presence of *Chlamydia* DNA by *Chlamydia*-specific PCR targeting the intergenic spacer region (IGS) between *Chlamydia* 16S and 23S rRNA genes [26] and using primers cIGS1f (5'-CAAG GTG AG GCTG ATG AC-3') and cIGS2r (5'-T CGC CTK TCA ATG CCA AG-3'). PCR conditions are described elsewhere [26]. The identity of all positively tested IGSP PCR products was determined by direct sequencing of the PCR product from both strands. Sequencing was performed with an ABI Prism 377D DNA sequencer (Applied Biosystems) or Applied Biosystems 3100 (Syngene Biotech). The obtained sequences were compared with the sequences available in GenBank using the BLAST server from the National Center for Biotechnology Information [27].

Investigation of abortion cases

Abortion cases in the flock were further investigated for the presence of *Chlamydia* by routine bacteriology and immunohistochemistry of the placenta and the fetuses (lung, liver, kidney) as described elsewhere [28].

Statistical analysis

Each ELISA antibody value was initially categorized into positive, questionable or negative as described previously [17,24]. For the analysis, questionable and negative results were both interpreted as negative. Whole flock response patterns over time were visualized using box plots. For the hypotheses tested, the first time, the proportion of positive cases to achieve time point was compared with the expected flocking of flocks. Exact tests with exact P -values. In addition, the means of the hypotheses were compared using a repeated measures ANOVA with animal ID, time (within animal repetition factor), vaccination status (flocks B and E only), and the interaction between time and vaccination (again only for flocks B and E).

Data were stored and handled in Microsoft Excel, and analysed using the statistical software packages NCSS2004 [29] and SPSS14 [30]. The overall level of statistical significance was set at 0.05.

Authors' contributions

AG carried out the serum sampling and the serological investigations and drafted the manuscript. R T performed the investigation of the abortion cases and contacted the flock owners. E V and E P prepared the cELISA plates. CK investigated the ewe samples by PCR. MGD performed the statistical analysis. D RZ performed the DNA sequencing. A P assisted in the drafting and editing of the manuscript. A P and N B participated in the design and coordination of the study. All authors read and approved the final manuscript.

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Figure legend

Figure 1 - Box plots of cELISA antibody values of all examined sheep over the four investigation dates.

Some of the animals infected with A, B and E were vaccinated at different times (gray boxes).

Tables

Table 1 - Serological results A, B, C, D and E

cELISA positive (above cutoff) sheep with frequency, respective proportion (%),
median and interquartile range.

Flock(n)	Parameter	Spring2005	Autumn2005	Spring2006	Autum2006
A	No. positive	15	13	11	11
15s heep	Prop. Pos.(%)	100	87	73	73
	Median titer	91.7	86.6	81.3	82.3
	Titerrange	70.9 –99.9	55.1–99.2	62.0–99.0	55.3–99.2
B	No. positive	8	4	2	8
26s heep	Prop. Pos.(%)	31	16	8	31
	Median titer	65.0	62.9	58.7	63.9
	Titerrange	61.3 –72.8	55.1–68.1	56.4–61.0	55.3–77.5
C	No. positive	14	13	14	15
17s heep	Prop. Pos. (%) ¹	82	76	82	88
	Median titer	82.9	72.2	71.3	76.8
	Titerrange	69.1 –95.2	55.1–89.6	57.1–94.8	55.4–95.8
D	No. positive	13	21	29	19
63s heep	Prop. Pos.(%)	21	33	46	30
	Median titer	69.5	69.1	69.4	74.3
	Titerrange	55.1 –100	55.1–93.4	55.6–88.6	56.9–95.2
E	No. positive	26	24	19	24
63s heep	Prop. Pos.(%)	42	38	30	38
	Median titer	73.4	81.2	80.4	81.8
	Titerrange	57.4 –94.8	56.5–96.3	55.2–97.6	56.0–98.7

¹significant difference in % positive (Fisher's Exact Test, p= 0.024)

Table 2 - Serological results vaccinated vs. non-vaccinated (Flock B and E)

Comparison of cELISA positive (above cut-off) vaccinated and naturally exposed sheep with frequency, respective proportion (%), median titer and titer range.

Flock(n)	Parameter	Spring2005	Autumn2005	Spring2006	Autum2006
B ¹	No. positive	6	3	2	2
14s heep	Prop. Pos.(%)	43	22	14	43
	Median titer	62.9	61.3	58.7	64.5
	Titer range	61.3 –64.2	55.1–66.3	56.4–61 .0	55.3–77.5
B ²	No. positive	2	1	0	2
12s heep	Prop. Pos.(%)	17	8	0	17
	Median titer	71.1	68.1	-	62.2
	Titer range	69.4 –72.8	-	-	55.9–68.5
E ¹	No. positive	1	0	0	1
13s heep	Prop. Pos.(%)	8	0	0	8
	Median titer	61.4	-	-	73.2
	Titer range	-	-	-	-
E ²	No. positive	25	24	19	23
50s heep	Prop. Pos.(%)	50	48	38	46
	Median titer	76.3	81.2	80.4	82.2
	Titer range	57.4 –94.7	56.5–96.3	55.2–97.6	56.0–98.7

¹vaccinated group

²non-vaccinated group

Table 3 - Flock details

Flock	Examination dates	Average no.s heep	Sheep tested all 4 times	Flock history	OEA status ¹	Vaccination with live vaccine
A	spring & autumn 2005/2006	54	15	chlamydial abortions in autumn 2004	positive	15s heep (spring 2005)
B	spring & autumn 2005/2006	48	26	chlamydial abortion outbreak in early 2003	negative	14s heep (spring 2003), no vaccination booster
C	spring & autumn 2005/2006	45 ²	17 ³	chlamydial abortions (positive control)	positive	no
D	spring & autumn 2005/2006	105	63	no abortions (negative control)	negative	no
E	spring & autumn 2005/2006	118	63	chlamydial abortions in the past	positive	13s heep (winter 2005)

¹OEA=ovine enzootic abortion

²Average no. goats: 11

³Goats tested all 4 times: 4

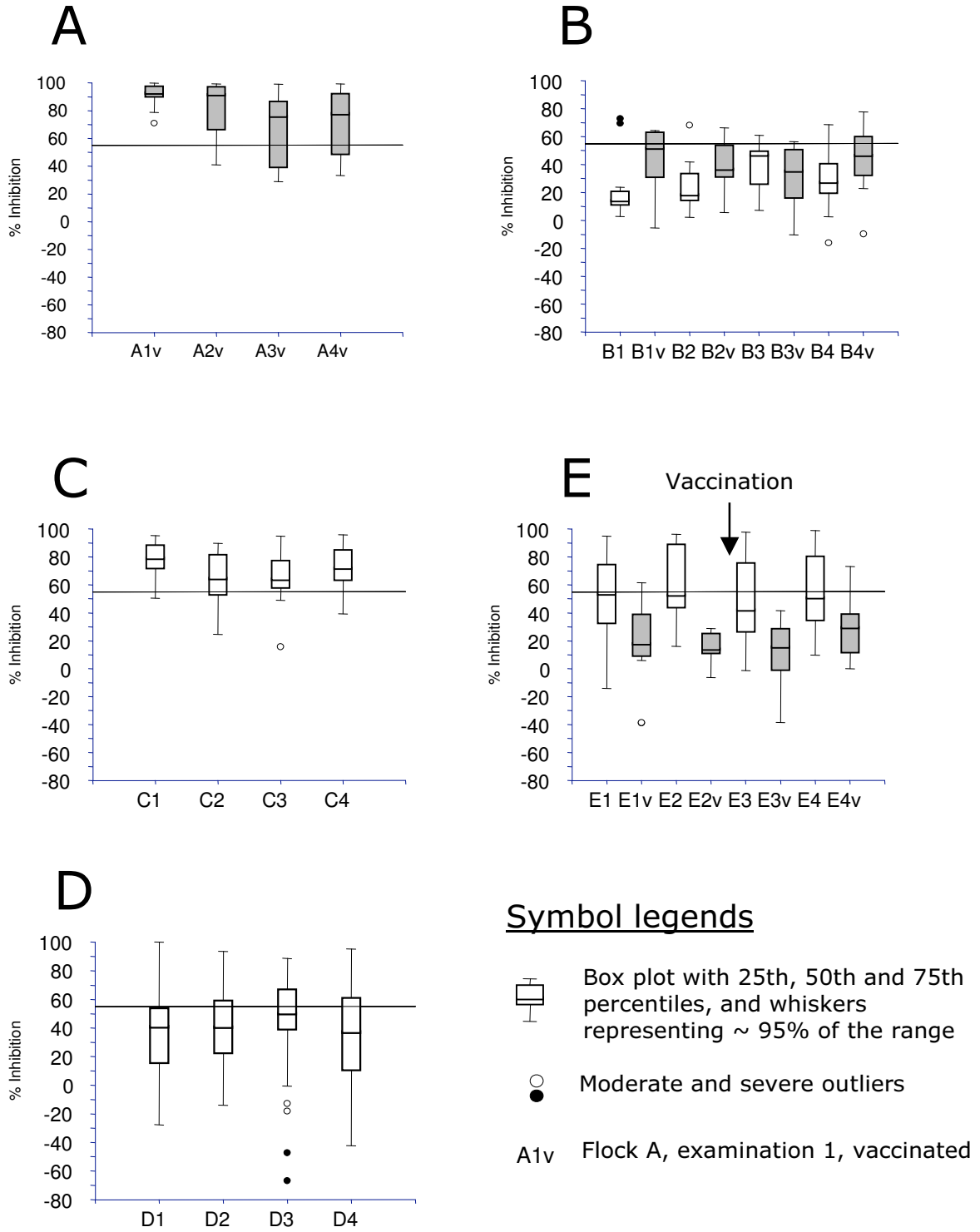


Figure 1